1 Supplemental material

2 Supplemental Figures



Supplemental Figure 1. FVIII staining specificity. Representative blots of FVIII-specific B cell staining performed on splenic cell suspensions from (A) naïve IgHEL mice (n=3) that express only HEL-specific B cell receptor, (B) HemA mice without (n=3) and (C) Hem A mice injected with 2 UI FVIII per mouse once per week (n=3) are shown.

PD-L1⁺ Tregs mediate immune tolerance in hemophilia



Supplemental Figure 2. PD-1 controls anti-FVIII antibody responses in FVIII-competent mice. 12 13 Experimental setup for A-H: Two UI/mouse of human rFVIII were intravenously injected into HemA 14 (red, n=5) and WT (black, n=4) in weekly intervals. One day after the last injection splenocytes were 15 further analyzed by flow cytometry. (A) Representative histograms of the PD-1 expression of FVIII-16 negative B cells. (B) Quantification of the percentage of PD-1 expressing FVIII-negative B cells. (C) 17 Molecular PD-1 expression on FVIII-negative B cells determined by flow cytometry. (D) Percentage of FVIII-specific B cells expressing Fas analyzed at day 22 ex vivo. (E) MFI of Fas expressing FVIII-18 19 specific B cells. (F) Quantification of the percentage of CD80 expressing FVIII-specific B cells. (G) 20 Molecular CD80 expression on FVIII-specific B cells. (H) Representative UMAPS of FVIII-specific B cells are shown to demonstrate PD-1 single expression or co-expression with CD80 or Fas. P values 21 were calculated using a Students T test. ns...not significant; *P <0.05. 22

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27 Supplemental Figure 3. CTLA-4 does not regulate inhibitor formation against FVIII. (A) Experimental scheme for B-H: WT mice were injected once a week with 2UI of rFVIII (black, n=6). 28 29 One group of WT mice additionally received an anti-CTLA-4 inhibitory antibody that was applied 30 twice a week (green, n=7) intraperitoneally. (B) Quantification of the number of FVIII-specific B cells 31 in the spleen of WT mice after treatment with or without anti-CTLA-4 by flow cytometry. (C) 32 Percentage of PD-1 expressing FVIII-specific B cells and (D) MFI of PD-1 on FVIII-specific B cells 33 at day 22. (E) Quantification of the percentage of apoptotic FVIII-specific B cells by analyzing the 34 expression of Annexin V and Hoechst. (F, G) ELISA-based quantification of the FVIII-specific IgG 35 antibody titer (F) and Bethesda units per ml (G) in the serum as well as the percentage of residual active FVIII (H) in the plasma with a COATEST assay in WT and WT mice treated with an anti-36 37 CTLA-4 blocking antibody. P values were calculated using a Students T test. ns...not significant P>0,05. 38



PD-L1⁺ Tregs mediate immune tolerance in hemophilia

41 Supplemental Figure 4. Enhanced PD-L1 expression on Tregs in WT mice. (A-S) Two UI/mouse 42 of human rFVIII were intravenously injected into HemA (red, n=6) and WT (black, n=5) in weekly 43 intervals. One day after the last injection splenocytes were further analyzed by flow cytometry. Within 44 each histogram, HemA mice are displayed in red and WT mice in grey in comparison to the Pd-l1 45 knockout control (black line). (A) Representative histograms of the PD-L1 expression on CD4⁺Foxp3⁺ 46 T cells of immunized mice and naïve HemA mice (light red) or WT control (grey). (B) Percentage of PD-L1 expression and (C) the amount of PD-L1 per cell on regulatory T cells gated as CD4⁺Foxp3⁺ 47 48 T cells. (D) Representative histograms of the PD-L1 expression on activated CD4⁺CD44⁺ T cells. (E) 49 Percentage of PD-L1 expression and (F) the amount of PD-L1 per cell on activated T cells gated as 50 CD4⁺CD44⁺ T cells. (G) Representative histograms of the PD-L1 expression on follicular T helper 51 cells. (H) Percentage of PD-L1 expression and (I) the amount of PD-L1 per cell on follicular T helper 52 cells gated as CD4⁺CXCR5⁺PD-1⁺T cells. (J) Representative histograms of the PD-L1 expression on 53 CD8 T cells. (K) Percentage of PD-L1 expression and (L) the amount of PD-L1 per cell on CD8 T 54 cells gated as CD8+ T cells. (M) Representative histograms of the PD-L1 expression on gd T cells. 55 (N) Percentage of PD-L1 expression and (O) the amount of PD-L1 per cell on gd T cells gated as 56 CD45⁺CD3e⁺gdTCR⁺ T cells. (P) Representative histograms of the PD-L1 expression on NKT cells. 57 (Q) Percentage of PD-L1 expression and (R) the amount of PD-L1 per cell on NKT cells gated as 58 $CD45^{+}CD3e^{+}NK1.1^{+}$ T cells. P values were calculated using a Students T test. *P <0.05; **P < 0.01. 59





Supplemental Figure 5. Treg depletion efficiency. In Foxp3-LuciDTR mice Foxp3⁺ (n=5) Tregs were depleted by injecting 15 ng/g mouse DTX intraperitoneally at day -1 and 0 of the experiment. Depletion efficiency was controlled in blood samples of DTX treated and untreated mice (n=3) by flow cytometry. Representative Dot blots and histograms are shown. Up to 97% of all eGFP (Foxp3)⁺ Tregs were depleted.



73 Supplemental Figure 6. No differential PD-L1 or PD-1 expression in naïve HemA and WT mice. Splenocytes of naïve HemA (n=5) and WT mice (n=5) were analyzed by flow cytometry. (A) 74 Determination of the number of Tregs gated as CD4⁺Foxp3⁺, as well as their percentage (B) and their 75 76 molecular expression of PD-L1 per cell (C). (D) Number of FVIII-specific Tregs gated as single living 77 B220⁻ CD4⁺ CD25⁺ CD127⁻ T cells. (E) Determination of the proportion of PD-L1⁺ FVIII-specific 78 Tregs. (F) PD-L1 expression per cell quantified as gMFI on FVIII-specific Tregs. (G) number of 79 FVIII-specific B cells analyzed by flow cytometry. (H) PD-1 expression shown as % and (I) MFI of 80 PD-1 on FVIII-specific B cells. P values were calculated using a Students T test. ns...not significant 81 P >0.05.



Supplemental Figure 7. PD-L1 competent Tregs reduce inhibitor titers in HemA mice. Tregs were isolated either from WT or PD-L1^{-/-} mice and 1x10⁶ cells were transferred into HemA mice by intravenous injections. Starting from the next day, HemA (red, n=9) and HemA mice that received Tregs from WT (blue, n=9) or PD-L1^{-/-} (purple, n=9) mice were intravenously injected with 2 UI/mouse of rFVIII in weekly intervals. One day after the last injection blood serum was harvested. Bethesda units per ml were measured by ELISA in serum of the indicated mice. P values were calculated using one-way ANOVA and Bonferroni posttest. *P <0.05.</p>

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Supplemental Figure 8. Treg-mediated suppression of antibody-producing B cells is PD-1 95 96 dependent. HemA mice (n=3) were immunized 3 times with 2UI of FVIII in a weekly interval. Afterwards, plasma cells were isolated from splenic cell suspensions and 3x10⁵ B cells were 97 cocultured with 5x10⁵ Tregs isolated from naïve WT mice for 16 hours with or without a PD-1 98 99 blocking antibody (RPM1-14; 40 µg/ml). (A) Percentage of early apoptotic B cells after co-culture. (B) overall percentage of surviving B cells after 16 hours of co-culture. (C and D) 0,5x10⁶ naïve 100 splenocytes from HemA mice (n=5) were co-cultured with either 0.5×10^6 wildtype Tregs or T helper 101 102 cells for 16 hours with or without FVIII protein. Quantification of the proportion of early apoptotic 103 FVIII-specific B cells (C) or FVIII-unspecific B cells (D). P values were calculated using one-way ANOVA and Bonferroni posttest. ns...not significant P >0.05, *P <0.05, *P <0.001. 104



106 Supplemental Figure 9. ITI-like treatment increases active FVIII levels but does not diminish 107 inhibitors in HemA mice. Experimental setup for A-C: 2 UI/mouse of human rFVIII were 108 intravenously injected into HemA (red, n=4) mice in weekly intervals for three weeks, and twice a 109 week for the high dose FVIII regime (blue, n=5). Compared to the therapeutic FVIII treatment (once 110 a week), the high dose FVIII application regime (twice a week) is used to induce tolerance (short ITI) 111 in HemA mice. One day after the last injection blood serum and plasma were harvested and further 112 analyzed by ELISA or COATEST assay, respectively. (A) Quantification of the FVIII-specific IgG 113 antibody titer and (B) Bethesda units per ml in the serum. (C) The percentage of residual active FVIII 114 was determined in the plasma of HemA mice treated with FVIII once or twice a week. P values were calculated using a Students T test. ns...not significant; **P < 0.01. 115

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Supplemental Figure 10. Foxp3 expression of sorted CD25⁺CD127⁻CD4⁺ T cells. Regulatory T cells of Foxp3-LuciDTR mice (n=4) were sorted either as CD4⁺eGFP⁺(Foxp3⁺) or as CD4⁺CD25⁺CD127⁻ T cells. As a control CD4 T helper cells were sorted as CD4⁺ eGFP⁻(Foxp3⁻) T cells. RNA isolation and a subsequent qRT-PCR was performed to determine the expression of *Foxp3* in these cells. Expression is correlated to one mean of Tregs sorted as CD4+eGFP+ T cells. P values were calculated using one-way ANOVA and Bonferroni posttest. ns...not significant P>0,05; ***P < 0.001.</p>

PD-L1⁺ Tregs mediate immune tolerance in hemophilia



132 Supplemental Figure 11. ITI-induced Tregs are mainly peripherally derived Tregs. (A) 2 133 UI/mouse of human rFVIII were intravenously injected into HemA mice (red, n=4) or WT mice 134 (black, n=5) in weekly intervals for 3 weeks, and twice a week for the high dose FVIII regime (blue, 135 n=5). At day 22, splenic suspensions were analyzed by flow cytometry to discriminate the proportion 136 of peripherally-derived (Neuropilin-1 negative) FVIII-specific Tregs. FVIII-specific Tregs were 137 identified as living, single B220-CD4+CD25+CD127- T cells. Error bars represent SEM. P values were calculated using one-way ANOVA and Bonferroni posttest. ns...not significant, **P < 0,001. 138 139 (B-E) 2 UI/mouse of human rFVIII were intravenously injected into HemA mice (red, n=4) in weekly 140 intervals for 3 weeks, and twice a week for the high dose FVIII regime (blue, n=6). At day 22, splenic 141 suspensions were analyzed by flow cytometry to determine the expression of the exhaustion markers 142 PD-1 (B), TIM-3 (C), LAG-3 (D) and CTLA-4 (E), per cell on living, single FVIII-specific 143 (tetramer+) CD4 T cells. P values were calculated using a Students T test. ns...not significant P>0,05. 144

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150 Supplemental Figure 12. Decreased Bcl-2 expression in FVIII-specific B cells of ITI-treated

151 **mice.** (A-C) Two UI/mouse of rFVIII were intravenously injected into HemA (red, n=4) mice in

152 weekly intervals. Immune tolerance induction in HemA mice (blue n=5) was achieved by injecting

- 153 of rFVIII twice a week. (A) Representative histograms of Bcl-2 expression in FVIII-specific B cells.
- 154 (B) Quantification of the proportion of Bcl-2 expressing FVIII-positive B cells. (C) Mean Bcl-2
- 155 expression per cell in FVIII-positive B cells. P values were calculated using a Students T test. *P <
- 156 0.05 (D) Two UI/mouse of rFVIII were intravenously injected into HemA (red, n=4) or WT (black,
- 157 n=5) mice in weekly intervals. Tolerized HemA mice (blue n=5) were injected twice a week with
- 158 rFVIII. Mean expression of IRF4 per cell in FVIII-positive B cells is displayed. P values were
- 159 calculated using one-way ANOVA and Bonferroni posttest. **P < 0,001
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Supplemental Figure 13. Inhibitor formation in HemA mice. Two UI/mouse of human rFVIII were intravenously injected into HemA mice (n=14) in weekly intervals. HemA mice were bled 14 days after initial injection to measure IgG antibodies (A) and inhibitor production (B) in the serum of the mice via ELISA. Based on these results the mice were equally distributed amongst all three group to obtain a comparable pre-treatment titer as depicted in A and B. P values were calculated using oneway ANOVA and Bonferroni posttest. ns...not significant P>0,05.

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PD-L1⁺ Tregs mediate immune tolerance in hemophilia



173 Supplemental Figure 14. Inhibitory ligand expression on FVIII-specific B cells in humans. (A-

174 C) mRNA was extracted from sorted FVIII-specific B cells of the patient on ITI shown in Figure 6B

175 (red) or healthy controls (black) and analyzed by RT-PCR at different time points during ITI. Relative

- 176 mRNA expression of PD-L1 (A), PD-L2 (B), and FASL (C) in FVIII-specific B cells. Expression is
- 177 correlated to one healthy individual. Data are presented as mean +/- SEM.
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FVIII-specific tetramer

Supplemental Figure 15. FVIII-specific tetramer staining of CD4 T cells in humans. Blood was collected from HLA-matched (HLA15.01, n=3) or -mismatched hemophilia A patients (n=3). The specificity of the tetramer staining was verified by flow cytometry. The cells were gated on living single cells and the binding capacity is displayed on CD4⁺T cells. Representative dot plots are shown.